



Protocol for Immunohistochemical use of Monoclonal Antibodies to Dystrophin; VP-D505, VP-D507 and VP-D508

1. Freeze muscle blocks in isopentane chilled in liquid nitrogen.
2. Cut 4-10 μ m sections and air dry on slides coated with section adhesive.
3. Slides may be stored at -70 °C wrapped in cling film until required. If stored sections are used, allow sections to equilibrate to room temperature before unwrapping and proceeding.
4. Apply primary antibody to section (unfixed). Incubate for 1 hour at 25 °C.
5. Wash sections 3 x 10 minutes in buffer.
6. Apply labeled secondary antibody (e.g. FI-2000 or PI-2000). Incubate for 1 hour at 25 °C.
7. Wash sections 3 x 10 minutes in buffer.
8. Mount fluorescent sections in VECTASHIELD® Mounting Medium (H-1000) or visualize peroxidase label (e.g. by exposure to freshly prepared DAB, SK-4100). Dehydrate, clear and mount peroxidase labeled sections for permanent preparations.

References

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