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**Protocol for immunohistochemical use of the following monoclonal antibodies:**

**VP-A102, VP-A105, VP-B205, VP-B206, VP-D501,  
VP-D503, VP-D504, VP-G803, VP-M648, VP-M664,  
VP-M665, VP-M666, VP-M667, VP-S283**

1. Freeze muscle blocks in isopentane chilled in liquid nitrogen.
2. Cut 4-10 $\mu$ m sections and air dry on slides coated with section adhesive.
3. Slides may be stored below -70 °C wrapped in cling film until required. If stored sections are used, allow sections to equilibrate to 25 °C before unwrapping and proceeding.
4. Apply primary antibody to section (**unfixed**). Incubate for 1 hour at 25 °C. Please note that where VP-D503 and VP-D504 primary antibodies are used, it is recommended that sections are fixed in acetone/methanol (1:1) for 4 minutes at room temperature prior to incubation with the primary antibody.
5. Wash sections 3 x 10 minutes in buffer.
6. Apply labeled secondary antibody (e.g. FI-2000 or PI-2000). Incubate for 1 hour at 25 °C.
7. Wash sections 3 x 10 minutes in buffer.
8. Mount fluorescent sections in VECTASHIELD® Mounting Medium (H-1000) or visualize peroxidase label (e.g. by exposure to freshly prepared DAB; SK-4100). Dehydrate, clear and mount peroxidase labeled sections for permanent preparations using VectaMount™ (H-5000).